# CORRELATIVE MICROSCOPY OF CEREBELLAR STELLATE NEURONS. A REVIEW 

Orlando J. Castejón.<br>Biological Research Institute"Drs. Orlando Castejón and Haydée Viloria de Castejón". Faculty of Medicine. Zulia University. Apartado 526. Maracaibo, Venezuela.<br>Corresponding autor, e-mail: ocastejo@.cantv.net, phone: 58-261-7414370, fax: 58-261-7831611.

Recibido: Agosto 2011. Aprobado: Noviembre 2011.
Publicado: Noviembre 2011.


#### Abstract

The stellate neurons of cerebellar cortex molecular layer are easily identified at light microscopy, scanning and transmission electron microscopy levels since they are the only short axon nerve cells existing in the middle and outer thirds of cerebellar cortex molecular layer. Some internal details of fractured stellate neurons, such as GERL complex, endoplasmic reticulum and nuclear chromatin have been three-dimensionally viewed by scanning electron microscopy and the freeze-fracture method, taking advantage of the washing out of cytoplasmic soluble proteins from the fracture face induced by the freeze-fracture process. Freeze-etching replica technique for transmission electron microscopy shows the distribution of nuclear pores and the three-dimensional relief of endoplasmic reticulum and cell organelles. The stellate cell axon appears as a unique process directed toward Purkinje secondary and tertiary dendritic branches. The contoured stellate dendrites exhibit a beaded aspect and frequent bifurcations. Parallel and climbing fibers, and basket cell axons establish axospinodendritic and axosomatic contacts with stellate neurons and their dendritic processes. Axodendritic connections between stellate cells also are distinguished. Stellate neurons exhibit immunopositive reactions for Synapsin-I, PSD-95, GluR1, CaMKII and N-cadherin.


Keywords: Stellate neuron, cerebellum, correlative microscopy, immunohistochemistry.

## MICROSCOPÍA CORRELATIVE DE LAS NEURONAS ESTRELLADAS CEREBELOSAS

## RESUMEN

Las neuronas estrelladas de la capa molecular del cerebelo son facilmente identificables al microscopio óptico, electrónico y de barrido por ser las únicas neuronas de axón corto existentes en el tercio medio y externo de la capa molecular de la corteza cerebelosa. Algunos detalles internos del citoplasma, tales como el complejo de Golgi, el retículo endoplasmático, y el núcleo se visualizan mediante microscopía elctrónica de barrido y la técnica de criofractura para microscopía de barrido, debido al lavado de las proteínas solubles del citoplasma por la técnica de criofractura. La técnica de crifractura para microscopía electrónica de transmisión mostró la distribución de los poros nucleares y el relieve tridimensional del retículo endoplasmático y los organelos celulares. El axón único de las neuronas estrelladas aparece dirigido hacia las ramificaciones dendríticas secundarias y terciarias de las células de Purkinje. Las dendritas muestran aspecto contorneado, aspecto varicoso y frecuentes bifurcaciones. Las fibras paralelas, trepadores y axones de células de cesta establecen sinapsis axosomáticas espinodendríticas asimétricas con las dendritas de las neuronas estrelladas. Tambien se observan sinapsis axodendríticas entre células estralladas. Las neuronas estrelladas muestran reacciones inmunopositivas para Sinapsina I, PSD-95, GluR1, CamKII y N- caderina.

Palabras claves: Neuronas estrelladas, cerebelo, microscopía correlativa,inmunohistoquímica

## INTRODUCTION

Stellate cells, the intrinsic interneurons of the cerebellar molecular layer, were earlier described at light microscopy level by Fusari [1], Ramón y Cajal P. [2], Ponti [3], Ramón y Cajal S. [4], Smirnow [5], Estable [6], Jakob [7], Scheibel and Scheibel [8] and Fox et al. [9]. The transmission electron microscopic (TEM) features were earlier studied by Herndon [10], Fox et al. [9], Lemkey-Johnston and Larramendi [11,12], Castejón [13], Sotelo [14,15], Palkovits et al. [16], and Mugnaini [17]. The most complete description of these
neurons has been given by Chan-Palay and Palay [18], and Palay and Chan-Palay [19] by means of camera lucid drawings of Golgi light microscopy preparations and transmission electron microscopy (TEM). Castejón and Castejón [20)], and Castejón [21-23] described the TEM features of stellate neurons of mouse cerebellar cortex. Later, Castejón and Castejón [24] reported the glycosaminoglycan content, freeze-etching features, and three-dimensional morphology of stellate neurons by scanning electron microscopy (SEM). Castejón et
al. [25], using SEM and TEM, described the axodendritic connections between granule cell axons or parallel fibers and stellate cell dendrites. Obata et al. [26] firstly described putative GABA-releasing terminals from basket/stellate and Golgi cells immunostained with glutamate decarboxylase-67 antibody. Benagiano et al. [27] demonstrated GABA immunoreactivity in the cell bodies of stellate neurons of human cerebellar cortex by light and electron microscopy. Biagotti et al. [28] found by means of electron microscopy analysis that the basket and stellate cells, as well as the Golgi cells, have a remarkable glucose-6-phosphatase deshydrogenase (G6PD) activity. Corticotropin-releasing factor and urocortin espression in stellate neurons were found by Swinny et al. [29]. Fritschy et al. [30] found GABAergic axodendritic synapses of stellate neurons on Purkinje cells. More recently, Astori et al. [31] have demonstrated GABA release from stellate neurons onto Purkinje cells. Castejón [32] recently described Synapsin-I, PSD-95, GluR1, N-cadherin and Ca ${ }^{2+}$ /Calmodulin-dependent Protein Kinase II Alpha immunopositive reactions of stellate neurons.

The present review describes the light and Golgi light microscopy, SEM and TEM features of stellate neurons, and positive immunohistochemical activity of Synapsin-I, PSD-95, GluR1, Calcium/calmodulindependent protein kinase II (CaMKII) of stellate neurons, and N -cadherin, and their synapses with granule cell axons, climbing fibers, and from axonal terminals of neighboring stellate neurons.

## LIGHT MICROSCOPY

Plastic semithin sections of mouse cerebellar cortex stained with toluidine blue show the triple layered structure of cerebellar cortex formed by granule cell, Purkinje cell and molecular layers. The distribution of stellate neurons can be appreciated in the middle and outer third molecular layer [13]. (Fig.1).


Fig. 1. Mouse cerebellar cortex showing the stellate neuron (SN) distribution in the molecular layer. Basket cells (BC), Bergmann glial cells (BG), Purkinje cells (PC), and the granule cell groups (GC) also are distinguished. (Castejón, 1968).

## Golgi light microscopy

Close examination of the molecular layer with Golgi light microscopy technique shows the topographic relationship of stellate neurons with the secondary and tertiary Purkinje dendritic ramifications. The stellate cell axons directed to the Purkinje dendritic processes extend in the middle and outer thirds of molecular layer (Fig. 2).


Fig. 2. Mouse cerebellar cortex. Golgi stained thick paraffin section showing a stellate neuron (SN) sending its axonal process (arrow) toward the Purkinje tertiary spiny dendrites (PD). Some stellate neuron dendrites also are seen spreading in the molecular layer (Castejón, 1968).

In addition, Paula-Barboza et al. [33] described by means of the combined Golgi light microscopy and

Golgi-ultrastructural method the stellate cell axonal descending branches forming a pericellular basket around Purkinje cell soma, and contributing to the pinceaux surrounding the Purkinje axonal initial segment, where they establish septate-like junctions.

## CONVENTIONAL SCANNING ELECTRON

 MICROSCOPYFish and human cerebellar cortex specimens conventionally processed for conventional SEM, and ethanol-cryofracturing tecnique [34-38] show the superficial short-axon stellate neurons with round, elliptical or fusiform somata in a parasaggital fracture of the outer third molecular layer. These superficial stellate cells are easy to recognize, since they are the only neurons in the upper molecular layer, and appear surrounded by bundles of passing granule cell axons or parallel fibers [39]. (Fig.3).


Fig. 3. Teleost fish cerebellar cortex. Scanning electron micrograph showing the stellate neuron (SN) in the outer third molecular layer surrounded by bundles of parallel fibers (PF). The arrow indicates the parallel fibers approaching to the stellate cell soma (Castejón, 1988).

Three to five beaded short and ramified dendrites radiate from the cell body toward the neighboring Purkinje dendrites or other stellate cells [36-38]. The axon originates by way of a typical triangular shaped axon hillock and, after a short initial segment bifurcates into tenuous varicose collaterals. The short axonal process directed to the Purkinje cell dendrites, and the
convoluted and cryodissected dendritic processes can be appreciated in SEM human cerebellar cortex prepared by means of ethano-cryofracturing tecnique of Humphreys et al. [35]. (Figs. 4 and 5).


Fig. 4. Scanning electron microscopy of human cerebellar cortex showing a stellate neuron (SN), and its axonal processes (arrows) directed toward the Purkinje dendrite (Pd). Gold-paladium coating. (Castejón and Castejón, 1987).


Fig. 5. Human cerebellar cortex. Conventional SEM and ethanol-criofracturing technique. Outer surface of a stellate neuron ( SN ) showing the dendritic processes (arrow) ending on dendritic twigs (arrowhead). (Castejón and Castejón, 1987).

## SEM and SEM freeze-fracture technique

By means of the freeze-fracture technique for SEM [25,35-40], the stellate neurons are fractured through the equatorial plane showing at low magnification the condensed pattern of nuclear heterochromatin, and the three-dimensional image of the GERL complex [41,42], formed by the Golgi cisternae and their sacs,
endoplasmic reticulum canaliculi, and lysosomes. The dendrites exhibit also the outer surface of endoplasmic reticulum. The inner cytoplasmic details have been visualized taking advantage of washing out of cytosol soluble proteins induced by the SEM freeze-fracture process [43,44]. (Fig.6).


Fig. 6. Teleost fish cerebellar molecular layer (ML) showing the fractured stellate neuron (SN). SEM freeze-fracture method. The nucleus ( N ), the GERL complex (arrowheads), and the endoplasmic reticulum (ER) are observed extending from the cell body to the dendritic process (arrow). (Castejón, 1988).

## TRANSMISSION ELECTRON MICROSCOPY

Stellate cell soma
The fine structure of stellate neurons shows the general features of a microneuron characterized by a scarce band of perinuclear cytoplasm containing rough endoplasmic reticulum caniliculi, free ribosomes and polysomes, bundles of microtubules, motochondria, Golgi complex, lysosomes, and coated vesicles [24]. (Fig.7).
In addition, Ruela et al. [45] earlier described cilia in stellate neurons of rat cerebellum. Castejón and Castejón [24] demonstrated by means of light and electron microscopy histochemistry the presence of a homogenous alcianophilic substance within the stellate neuron cytoplasm characterized as a glycosaminoglycan sensitive to hyaluronidase treatment, mainly hyaluronic acid.


Fig.7. Mouse cerebellar cortex. Transmission electron micrograph of stellate neuron showing the nucleus ( N ), the cytoplasm containing scarce profiles of endoplasmic reticulum, numerous free ribosomes, bundles of microtubules ( Mt ), and mitochondria (M). The neighboring neuropil (NL) of molecular layer also is noted. (Castejón and Castejón, 1987).

Monteiro [46] established by means of a morphometric analysis at TEM level statistically significant differences in data concerning perikaryon volume, perikaryon surface and intracellular organelle composition between basket and stellate cells, and postulated that each class of interneuron should be designated with a specific name.

A complex neuropil formed by the Purkinje-parallel and climbing fiber spine synapses, surrounded by the Bergmann glial cell cytoplasm is observed adjacent to the stellate neurons.

## Stellate neuron synapses

According to earlier TEM studies, climbing and parallel fibers, and basket cell axons establish axosomatic and axodendritic contacts with stellate cells [11,19,24,39,47,48]. Small synaptic buttons of climbing fibers, "en passant" parallel fibers, basket cell and Lugaro cell axons, or axonic terminals of neighboring stellate neurons are observed attached to the somatic neuronal surface. Basket cell endings exhibit ellipsoidal and flattened synaptic vesicles. On the contrary, large synaptic endings of climbing fiber characterized by the presence of spheroid synaptic vesicles are observed making axodendritic junctions [49,50]. (Figs. 8 and 9).


Fig. 8. Mouse cerebellar molecular layer. TEM of stellate neuron displaying the nucleus ( N ), the mitochondria (M), Golgi complex (GC), scarce profiles of rough endoplasmic reticulum (ER), and an axosomatic synapse (AS, arrow). (Castejón and Castejón, 1987).


Fig. 9. Mose cerebellar cortex. Stellate neuronal soma (SN) exhibiting axosomatic synapses with a large climbing fiber synaptic ending (CF) characterized by spheroid synaptic vesicles, and with a small basket cell ending (BC) featured by flattened vesicles.

## TEM and the freeze-etching technique

With the freeze-etching technique the smooth fractured cytoplasm of stellate cells is observed exhibiting the three-dimensional relief of endoplasmic reticulum and cell organelles embedded in a smooth surface cytosol. Axodendritic contacts of climbing fibers also are found (Fig. 10).


Fig. 10. Mouse cerebellar molecular layer. TEM and freeze-etching technique. Stellate neuron cytoplasm (SN) displaying the endoplasmic reticulum profiles, free ribosomes embedded in the cytoplasm, and a dendritic process (D). A large climbing fiber synaptic ending (CF) appears synaptically apposed to the dendritic process (Castejón et al., 2000).

## CONFOCAL LASER SCANNING MICROSCOPY

## AND IMMUNOHISTOCHEMISTRY

Synapsin-I immunohistochemistry
Slices of rat cerebellar cortex labeled with Synapsin-I show the distribution of small puncta surrounding the stellate neurons and their process corresponding to the distribution of presynaptic endings of climbing fiber and parallel fibers [32,51]. (Fig. 11).

## PSD-95 immunohistochemistry

Slices of rat cerebellar cortex labeled with PSD-95 show the immunopositive stellate neurons and their process corresponding to the distribution of postynaptic endings of climbing fiber and parallel fibers [32,51]. (Fig. 12).
PSD-95 is a postsynaptic protein predominantly associated with postsynaptic densities [51].


Fig.11. Rat cerebellar cortex. Note the small puncta expressing Synapsin-I positive immunolabeling surrounding the stellate cells (SC) at the molecular layer. The Purkinje cell layer (PC) and granular layer (GL) also are appreciated. (Castejón and Dailey, 2009).


Fig.12. Rat cerebellar cortex immunolabeled with PSD-95 showing at the level of molecular layer the immunopositive stellate neuron ( SN ) corresponding to their postsynaptic axosomatic and axodendritic sites. Note also the immunopositive postsynaptic sites of granule cell dendritic tips at the glomerular regions $(\mathrm{GR})$ in the granular layer.

## GluR1 subunit immunohistochemistry

Rat cerebellar slices labeled with PSD-95 show small red puncta corresponding to GluR1 subunits of AMPA receptors surrounding stellate and basket cells in the rat molecular layer [32,52]. (Fig. 12).


Fig.13. High magnification of rat cerebellar slice double labeled with anti-GluR1 monoclonal antibody. The secondary antibody used was Alexa-488 goat antirabbit (GAR) IgG. Note the GluR1 immunoreactivity of stellate cell soma (SC) and processes, basket cell (BC), and Purkinje cell body (PC). (Castejón, 2010).

The distribution of GluR1 subunits relates this postsynaptic AMPA receptor subclass to the excitatory circuits of the cerebellar cortex formed by climbing and parallel fiber synapses upon stellate neurons. Thus far, the expression of another subunit, the GluR2 subunitscontaining AMPA receptors has been reported only on cerebellar stellate cells $[53,54]$.

## $N$-cadherin immunohistochemistry

Rat cerebellar slices double labeled with a primary antibody against N -cadherin, and Alexa 488 goat anti mouse (GAM)-antibody [32] show strong punctate immunostaining at the level of soma and processes of stellate neurons (Fig. 14).


Fig.14. Rat cerebellar slice. N-cadherin immunostaining showing immunopositive reaction of soma and processes of a stellate cell (SC), and Purkinje dendritic ramifications (PD). (Castejón, 2010).

N -cadherin is a membrane glycoprotein mediating strong homophilic adhesion and concentrated at the synaptic junctions and neural circuits, where they exert an active role in synaptic structure, function, plasticity, and in selective interneuronal connections during network function [55-60],


Fig.15. Rat cerebellar slice double labeled with double labeled with a primary antibody against CaMKII alpha, and a secondary antibody the Alexa 488 goat anti mouse (GAM)-antibody. Note the immunopositivity of stellate, basket and Purkinje cells. Immunohistochemistry of $\mathrm{Ca}^{2+} /$ Calmodulin-dependent Protein Kinase II Alpha

Stellate cell cytoplasm and processes show strong immunopositive reaction for $\mathrm{Ca}^{2+} /$ Calmodulindependent Protein Kinase II Alpha [32]. ( Fig. 15).

Calcium/calmodulin-dependent protein kinase II (CaMKII) is a $\mathrm{Ca}^{2+}$ - activated enzyme that is highly abundant in the brain and play a major role in $\mathrm{Ca}^{2+}-$ mediated signal transduction. CaMKII constitute a family of multifunctional protein kinase isoforms (alpha, beta, gamma and delta).[61,62]. Lisman et al. [62] have postulated their role in long-term information storage, motor learning and long-term synaptic memory.

## CONCLUSIONS

Correlative microscopy of stellate neurons made by light microscopy, transmission electron microscopy and freeze-etching technique, scanning electron microscopy and cryofracture method, as wells as the use of immunohistochemical techniques for confocal laser scanning microscopy have permitted a better and deeper understanding of cerebellar structure and function, mainly regarding the three-dimensional morphology of outer neuronal surface, intramembrane morphology, stellate cell synaptic axospinodendritic and axosomatic contacts with parallel and climbing fibers, and the precise localization of pre- and postsynaptic receptors by means of immunohistochemical techniques for Synapsin-I, PSD95, and GluR1 subtype of AMPA receptors. Stellate neuron synaptic contacts with Purkinje cell demonstrate the neural correlates of the inhibitory action of stellate cells upon Purkinje cells. The axosomatic and axodendritic presynaptic contacts of climbing and parallel fibers as demonstrated by transmission electron microscopy and Synapsin-I and PSD-95 evinces de excitatory action of these afferent fibers upon stellate neurons. The GluR1 immunopositivity of stellate neurons reveals the presence of glutamatergic neurotransmission at the level of stellate neuron synapses. The CaMKII immunopositivity of stellate neurons suggest their role
in long-term information storage, motor learning and long-term synaptic memory. The N -cadherin positive immunoreaction of stellate nerve cells and their intracortical circuits demonstrate the active role of these cell adhesion molecules in synaptic structure, function, and plasticity.

## ACKNOWLEDGEMENT

This review has been carried out by a subvention obtained from CONDES-LUZ and Fundadesarrollo.

## REFERENCES

[1]. Fusari R. (1983) "Sull órigine delle fibre nervose nello strato molecolare delle circonvoluzione cerebellari dell'uomo". Atiérrale Acad Sci (Torino) 19: 47-51.
[2]. Ramón y Cajal P. (1896) Las células estrelladas de la capa molecular del cerebelo de los reptiles. Rev Trim Micrograf 1: 149-150.
[3]. Ponti U. (1897) "Sull corteccia cerebellare della Cavia". Monit Zool Ital 8: 36-40.
[4]. Ramón y Cajal S. (1911) "Cellules éttoilée externes ou à cylindre-axe court". Histologie $d u$ Système Nerveux de l'Homme et des Vertébrés. Vol. 2. Paris: Maloine. Consejo Superior de Investigaciones Científicas. Madrid. Editor. pp 3032.
[5]. Smirnow R. (1897) "Uber eine besondere Art von Nervenzellen der Molecularschicht des Kleinhirns bei erwachsenen Saugetieren und beim" Menschen. Anat Anz 13: 636-642.
[6]. Estable C. (1923) "Notes sur la structure comparative de l'écorce cérébelleuse, et derivées physiologiques possibles" Trabaj Lab Invest Biol Univ Mad 21: 169-265.
[7]. Jakob A. (1928). "Das Kleinhirn". In: Mollendorff MV, editor. Handbuch der mikroskopischen anatomie des Menschen IV. Berlin: Julius Springer. pp 771-831.
[8]. Scheibel ME, Scheibel AB. (1954) "Observations on the intracortical relations of the climging fibers of the cerebellum". J Comp Neurol 101: 733-763.
[9]. Fox CA, Hilman HE, Siegesmund KA, Duta CR. (1967). "The primate cerebellar cortex. A Golgi and electron microscopic study". New York. Fox CA, Snider, RS, editors. Prog Brain Res 25: 175225.
[10]. Herndon RM. (1964) "The fine structure of rat cerebellum. II. The stellate neurons, granule cells, and glia". J. Cell Biol 23: 277-293.
[11]. Lemkey-Johnston N, Larramendi LMH. (1968a). "Morphological characteristics of mouse stellate, basket cells and their neuroglial envelope: an electron microscopic study". J Comp Neurol 134: 39-72.
[12]. Lemkey-Johnston N, Larramendi LMH. (1968b). "Types and distribution of synapses upon basket and stellate cells of the mouse cerebellum". $J$ Comp Neurol 134: 73-113.
[13]. Castejón OJ. (1968). "Electron microscopic observations at the level of cerebellar molecular layer. Doctoral Thesis". Invest Clin 27: 57-108.
[14]. Sotelo C. (1969) "Ultrastructural aspects of the cerebellar cortex of the frog".. Neurobiology of Cerebellar Evolution and Development. AMAERF Chicago Llinas R , editor. Institute for Biomedical Research. pp 327-371.
[15]. Sotelo C. (1970) "Stellate cells and their synapses on Purkinje cells in the cerebellum of the frog". Brain Res 17: 510-514.
[16]. Palkovits M, Magyar P, Szentagothai J. (1972) "Quantitative histological analysis of the cerebellar cortex in the cat. II. Structural organization of the molecular layer". Brain Res 34: 1-18.
[17]. Mugnaini E. (1972) "The histology and cytology of the cerebellar cortex.. The Comparative Anatomy and Histology of the Cerebellum. The Human Cerebellum, Cerebellar Connections and Cerebellar Cortex". Minneapolis. Larsell O, Jansen J, editors: The University of Minnesota Press. pp 201-251.
[18]. Chan-Palay V, Palay SL. (1972). "The stellate cells of the rat's cerebellar cortex". 136: 224-249.
[19]. Palay SL, Chan-Palay V. (1974). "Stellate cells". Cerebellar Cortex. Cytology and Organization. Berlin: Springer-Verlag. pp 216-233.
[20]. Castejón OJ, Castejón HV. (1981). "Transmission and scanning electron microscopy and ultracytochemistry of vertebrate and human cerebellar cortex". Glial and Neuronal Cell Biology. New York. Fedoroff S. editor A Liss. pp 249-258.
[21]. Castejón OJ. (1981) "Light microscope, SEM and TEM study of fish cerebellar granule cells". Scanning Electron Microsc IV: 105-113.
[22]. Castejón OJ. (1983). "Scanning electron microscope recognition of intracortical climbing fiber pathways in the cerebellar cortex". Scanning Electron Microsc III: 1427-1434.
[23]. Castejón OJ. (1984). "Low resolution scanning electron microscopy of cerebellar neurons and neuroglial cells of the granular layer". Scanning Electron Microsc III: 1391-1400.
[24]. Castejón, O.J., Castejón, H.V. (1987) "Electron microscopy and glycosaminoglycan histochemistry of cerebellar stellate neurons". Scanning Microscopy 1: 681-693.
[25]. Castejón OJ, Castejón HV, Apkarian RP. (2001). "Confocal laser scanning, conventional scanning and transmission electron microscopy of vertebrate cerebellar granule cells". Biocell 25: 235-255.
[26]. Obata K, Fukuda T, Konishi S, FY, Mitoma H, Kosaka T. (1999). Synaptic localization of the 67,000 mol. wt isoform of glutamate decarboxylase and transmitter function of GABA in the mouse cerebellum lacking the $65,000 \mathrm{~mol}$. wt isoform. Neuroscience 93: 1475-1482.
[27]. Benagiano V, Roncali L, Virgintino D, Flace P, Errede M, Rizzi A, Girolamo F, Robertson D, Bormann J, Ambrosi G. (2001). "GABA immunoreactivity in the human cerebellar cortex: a light and electron microscopical study". Histochem J 33: 537-543.
[28] Biagiotti E, Guidi L, Del Grande P, Ninfali P. (2003) "Glucose-6-phosphate dehydrogenase expression associated with NADPH-dependent reactions in cerebellar neurons". Cerebellum 2: 178-183.
[29]. Swinny JD, Metzger F, Ikema-Paassen J, Gounko NV, Gramsbergen A, Van Der Want JJ. (2004) "Corticotropin-releasing factor and urocortin differentially modulate rat Purkinje cell dendritic outgrowth and differentiation in vitro". Eur $J$ Neurosci 19: 1749-1758.
[30]. Fritschy JM, Panzanelli P, Kralic JE, Vogt KE, Sassoè-Pognetto M. (2006) "Differential dependence of axo-dendritic and axo-somatic GABAergic synapses on GABAA receptors containing the alpha1 subunit in Purkinje cells". $J$ Neurosci 26: 3245-3255.
[31]. Astori S, Luján R, Köhr G. (2009) "GABA release from cerebellar stellate cells is developmentally regulated by presynaptic GABA(B) receptors in a target-cell-specific manner". Eur J Neurosci 30: 551-559.
[32]. Castejón OJ. (2010) "Stellate neurons". Comparative and Correlative Microscopy of Cerebellar Cortex. Maracaibo. Venezuela. Astrodata. pp 170-185.
[33]. Paula-Barboza MM, Tavares MA, Rueda C, Barroca H. (1983) "The distribution of stellate cell descending axons in the rat cerebellum: a Golgi and combined Golgi-electron microscopical study". J Anat 137: 757-764.
[34]. Anderson RF. (1951) ""Techniques for the preservation of the three-dimensional structure in preparing specimens for the electron microscope. Trans Acad Sci (NewYork). 13: 130-134.
[35]. Humphrey WJ, Spurlock BO, Johnstons JS. (1974) "Critical point drying of ethonal-infiltrated cryfracture biological specimens for scanning electron microscopy". Scanning Electron Microsc I: 276-282.
[36]. Castejón OJ, Caraballo A. $\left(1980^{a}\right)$. "Application of cryofracture and SEM to the study of human
cerebellar cortex". Scanning Electron Microsc IV: 197-207.
[37]. Castejón OJ, Caraballo A. (1980b) "Light and scanning electron microscopic study of cerebellar cortex of teleost fish". Cell Tissue Res 207: 211226.
[38). Castejón OJ, Valero C. (1980) "Scanning electron microscopy of human cerebellar cortex". Cell Tissue Res 212: 362-374.
[39]. Castejón OJ. (1988) "Scanning electron microscopy of vertebrate cerebellar cortex". Scanning Microscopy II: 569-597.
[40]. Castejón OJ. (2003) "Stellate Cells". Scanning Electron Microscopy of Cerebellar Cortex. New York. Kluwer Academic/ Plenum Publisher. pp 81-85.
[41]. Novikoff AB. (1967) "Enzyme localization and ultrastructure of neurons". The Neuron. New York. Hyden G (Ed.). Elsevier. pp 255-318.
[42]. Novikoff AB. (1976). "The endoplasmic reticulum. A cytochemist's view (a review)". 73: 2781-2784.
[43]. Haggis GH, Bond EF, Phipps-Todd B. (1976) "Visualization of mitochondrial cristae and nuclear chromatin by SEM". Scanning Electron Microsc I: 282-286.
[44]. Haggis GH, Bond EF, Phipps-Todd B. (1977) "Freeze-fracture for scanning electron microscopy". J Microscopy 111:193-201.
[45]. Ruela C. Tavares MA, Paula Barboza MM. (1981) "Cilia in stellate neurons of the rat cerebellum". Experientia 37: 197-198.
[46]. Monteiro, RA. (1989) "Morphometric differences between basket cells and stellate cells of rat neocerebellum (Crus I and Crus II)". $J$ Submicrosc Cytol Pathol 21: 725-736.
[47]. Hamori J, Szentagothai J (1965) "The Purkinje cell baskets: Ultrastructure of an inhibitory synapse". Acta Biol Hung 15: 465-479.
[48]. Hamori J, Szentagothai J. (1966) "Identification under the electron microscope of climbing fibers and their synaptic contacts". Exp Brain Res 1: 65-81
[49]. Castejón OJ, Castejón, HV. (2001) "Correlative microscopy of cerebellar basket cells". J Submicros Cytol Pathol 33: 23-32.
[50]. Castejón OJ, Castejón HV, Alvarado MV. (2000). "Further observations on cerebellar climbing fibers. A study by means of light microscopy, confocal laser scanning microscopy and scanning and transmission electron microscopy". Biocell 24: 197-212.
[51]. Castejón OJ, Leah F, Dailey M. (2004). "Localization of Synapsin-I and PSD-95 in developing postnatal rat cerebellar cortex". Develop Brain Res 151: 25-32.
[52]. Castejón OJ, Dailey M. (2009) "Immunohistochemistry of GluR1 subunits of AMPA receptors of rat cerebellar nerve cells". Biocell 33:71-80.
[53]. Liu SJ, Cull-Candy SG. (2000) "Synaptic activity at calcium-permeable AMPA receptors induces a switch in receptor subtype". Nature 405: 454458.
[54]. Liu SJ, Cull-Candy SG. (2002) "Activitydependent change in AMPA receptor properties in cerebellum stellate cells". J Neurosci 22: 38813889.
[55]. Redies C. (1997) "Cadherins and the formation of neural circuitry in the vertebrate CNS". Cell Tissue Res 290: 405-413.
[56]. Redies C. (2000) "Cadherins in the central nervous system". Prog Neurobiol 61: 611-648.
[57]. Suzuki S, Sano K, Tanihara H. (1991) "Diversity of the cadherin family: evidence for eight new cadherins in nervous tissue". Cell Regul 2: 261270.
[58]. Suzuki SC, Inoue T, Kimura Y, Tanaka T, Takeichi M. (1997) "Neuronal circuits are subdivided by differential expression of type-I classic cadherins in postnatal mouse brains". Mol Cell Neurosci 9: 433-447.
[59]. Huntley GW, Benson DL. (1999) "Neural (N)cadherin at developing thalamocortical synapses provides an adhesion mechanism for the
formation of somatopically organized conecctions". J Comp Neurol 407: 453-471.
[60]. Huntley GW, Gil O, Bozdagi O. (2002) "The cadherin family of cell adhesion molecules: multiple roles in synaptic plasticity". Neuroscientist 8: 221-233.
[61]. Chang BH Mukherji S, Soderling TR. (2001) "Calcium/calmodulin-dependent protein kinase II
inhibitor protein: localization of isoforms in rat brain". Neuroscience 102: 767-777.
[62]. Lisman, J., Schulman H,Cline H. (2002) "The molecular basis of CaMKII. Function in synaptic and behavioural memory". Nature Rev Neurosci 13: 175-190.

